

Perfusion Fixation of Whole Mouse

1. Use 20cc of cold 0.9% saline
2. Use 40-60cc cold 3% glutaraldehyde (EM grade) in Sorensen's buffer
3. Place 20cc syringe on butterfly
4. Anesthetize (the Schimdt lab uses isofluorane)
5. **IN FUME HOOD with CHUX underneath**, Cut mouse up the midline and open the chest
6. Use hemostat to hold the skin back
7. Insert needle just left of the apex
8. Clip the atria
9. Push the saline through slowly
10. When the liver clears of blood, that is enough saline
11. Change to the glutaraldehyde syringe (60cc syringe)
12. While pushing on the syringe, hold butterfly down as well as the right leg of the mouse
13. Mouse will become stiff when finished
14. Rinse mouse before dissection with water (optional)
15. Harvest tissue right away and place in fixative
16. Flush butterfly with saline before the next mouse

Materials:

25g3/4 blood collection butterfly needle, VWR, VT7255, pack of 50

60cc syringe, VWR, 82002-318